

Drug Screening by Paper Spray Mass

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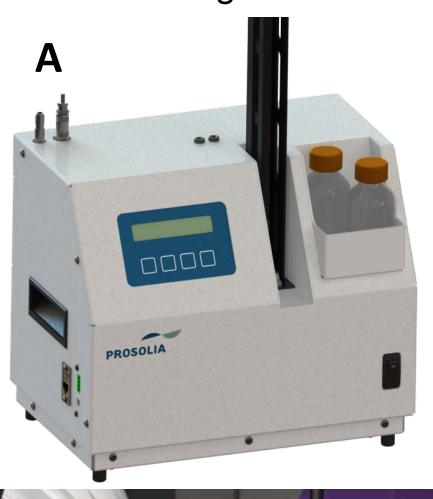
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Overview

- ☐ Paper Spray Mass Spectrometry is a method for direct analysis of biofluid samples, including dried blood spots. No sample preparation is needed
- ☐ Goal: develop a **drug screening** method for post-mortem forensic toxicology
 - Screen for 154 commonly encountered drugs and drug metabolites, including amphetamines, analgesics, anesthetics, anticonvulsants, antidepressants, antipsychotics, barbiturates, benzodiazepines, and opiates.
- ☐ A representative subset of the more challenging targets was selected for initial assessment and method development

Introduction

- ☐ An MS front-end is available that attaches to MS in place of the HPLC system and commercial electrospray/APCI source
- □ ~10 μL of sample is applied to a single-use cartridge containing the paper substrate
- ☐ An extraction/spray solvent is applied to the cartridge
 - > Solvent wicks through the paper and performs an extraction from the dried biofluid sample
 - > Electrospray ionization is induced directly from the paper tip
- ☐ Advantages: no chromatography, no sample preparation, small sample volume, no carry-over, no solvent waste, low solvent consumption
- ☐ Disadvantages: lower selectivity (no chromatography), higher detection limits



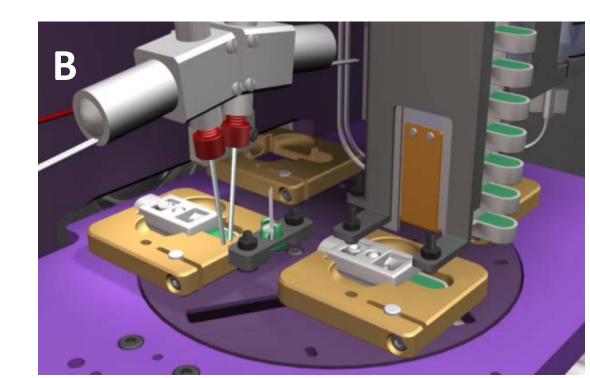
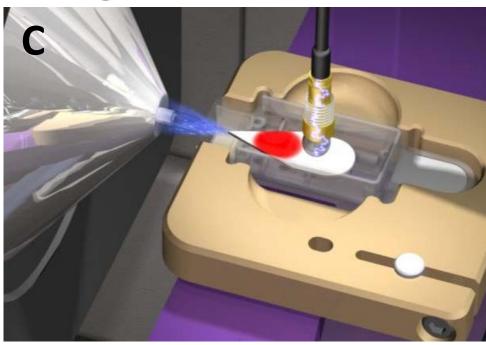


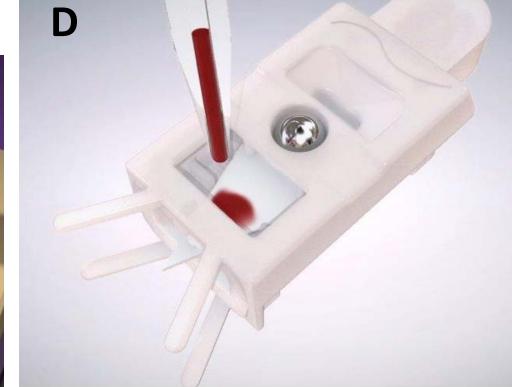
Figure 1.
A.Paper spray autosampler (Velox 360, Prosolia Inc.)

B.Inside autosampler - moving cartridges between stations



D.Paper spray cartridge with blood sample applied







Bottom: Paper after spray voltage is applied. Taylor cone is visible

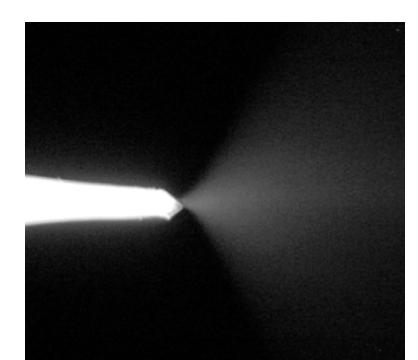
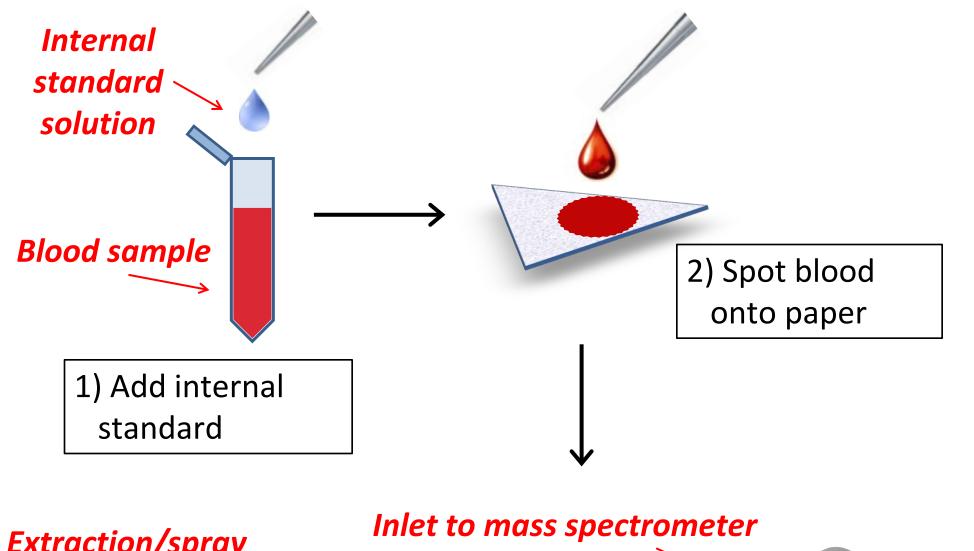


Figure 3. Picture of cone-jet generated from paper

Methods



Extraction/spray solvent (~100 μL) 3.5 kV Charged droplets

Sample loading

10-20 μ L of blood sample was pipetted onto the sample and allowed to dry. The blood sample can be mixed with an internal standard solution prior to spotting if desired.

Extraction and ionization

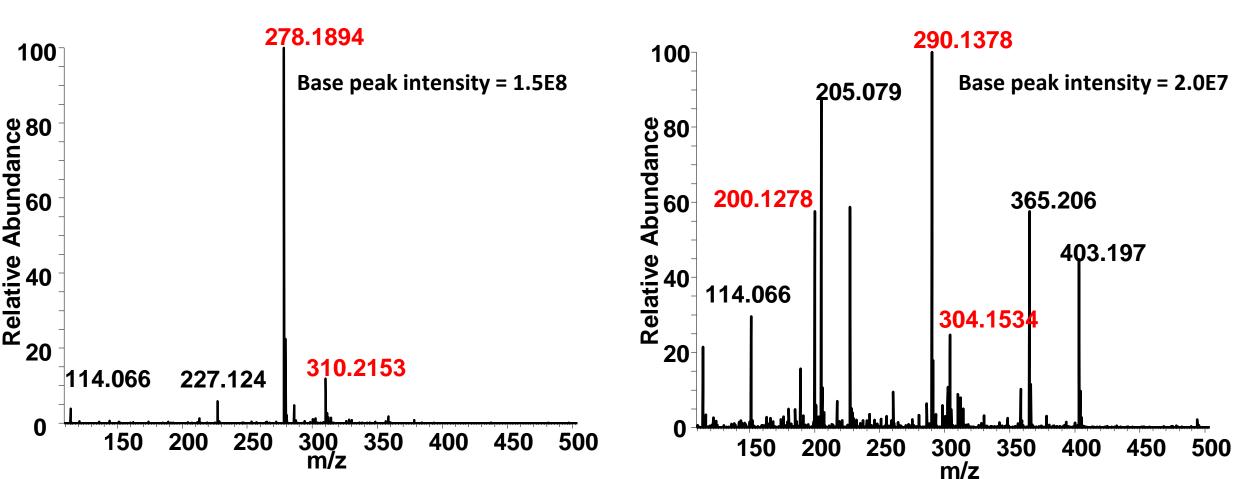
~100µL solvent is applied to the back of the paper. The solvent acts as both the extraction solvent and the electrospray solvent. Typical solvents include methanol or acetonitrile mixed with water (<10%) and acetic acid (<0.1%). Extraction step takes about 60 seconds.

Detection

The cartridge is positioned about 5mm in front of the inlet to the MS. Spray voltage of 3-5 kV is applied to the paper. Analyte signal is normally seen immediately. Signal duration is about 60 seconds.

Results

1. Drug Screening in Urine – Exact Mass on the Exactive



| Compound | Theoretical m/z (M+H) ⁺ |
|-----------------------|---------------------------------------|
| Ecgonine methyl ester | 200.1281 |
| Benzoylecgonine | 290.1387 |
| Cocaine | 304.1543 |
| EDDP | 278.1903 |
| Methadone | 310.2165 |

Figure 4. Two spectra obtained from paper spray MS of urine samples obtained from a methadone clinic. Analysis was performed on a Thermo Exactive MS

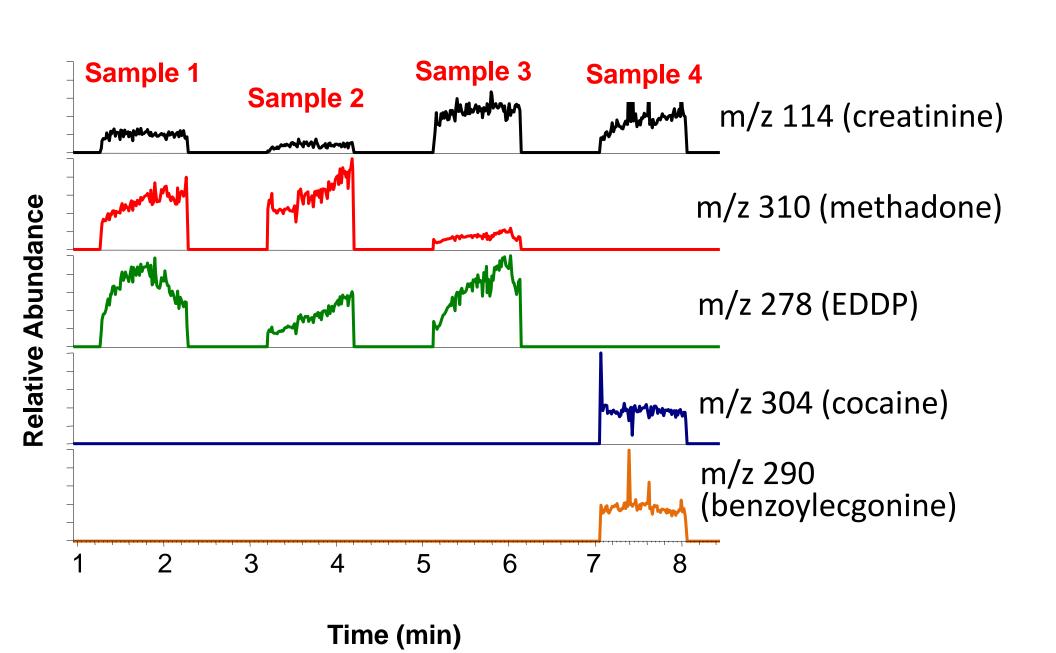


Figure 5. Extracted ion chronograms (5ppm window) for several compounds detected from the urine. Four samples are shown on each chronogram

lass Spectrometry: a Direct Analysis Approach

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Drug Screening in Blood – MS/MS on a Triple Quad

Table 1. Results for a representative subset of challenging drugs analyzed from human whole blood ☐ Challenging drugs were selected for initial assessment

Poor ionizers or low cutoff levels

□Screening cutoffs were set by consultation with AIT Laboratories

□Potential interferences with the same nominal mass were obtained from drugbank.ca and hmdb.ca (drug origin only)

| Drug | Target Cutoff (ng/mL) | SRM Transition | Signal to Blank at Cutoff | Potential Interferences (HMDB.ca) | Excluded Interferences |
|-------------------|-----------------------|-------------------|---------------------------------|--|---|
| | 5 | 309→281 | 8 | Pinazepam | Oxybuprocaine ¹ |
| Alprazolam | | 309→205 | 8 | Fluoxetine glucuronide 8-Hydroxycarteolol | Phenylbutazone ² Indecainide ³ |
| | 10 | 286→121 | 11 | Morphine Same as morphine | Same as morphine |
| 7-Aminoclonazepam | | 286→94 | 6 | | |
| A 1 | 50 | 136→91 | 64 | <u>Adenine</u> (136→119) | Homocysteine ¹ |
| Amphetamine | | 136→119 | 31 | | |
| Dunnananahina | 1 | 468→414 | 2 | 4-Hydroxytamoxifen sulfate | Tobramycin ³ Tiropramide ³ |
| Buprenorphine | | 468→396 | 2 | | |
| Clonazepam | 10 | 316→214 | 3 | Codeine N-oxide Rotigotine Saxagliptin | Bromazepam ¹ Efavirenz ¹ Alizapride ¹ Mitiglinide ³ Chlorprothixene ³ |
| Cionazepani | 10 | 316→241 | 5 | | |
| Cocaethylene | 50 | 318→196 | 68 | beta-oxycodol N2-Monodes-methylnizatidine Arbutamine | Nilutamide ² Nateglinide ² Tetrabenazine ³ Butenafine ¹ Acebutolol ¹ Berberine ¹ Acetyl-α-methylfentanyl ³ N-Desmethyl rosuvastatin ¹ |
| - Codacanyiene | | 318→82 | 51 | | |
| Fentanyl | 1 | 337→105 | 2 | Captopril-cysteine disulfide | |
| • | | 337→188 | 6 | | |
| Gabapentin | 500 | 172→137 | 44 | N.F. | Metronidazole ¹ Rasagiline ¹ |
| Gabapentin | | 172→119 | 48 | | |
| Ketamine | 100 | 238→125 | 48 | 2-Amino-5-benzoylbenzimidazole | N.F. |
| Retainine | | 238→89 | 37 | | |
| Meprobamate | 2000 | 219→180 | 67 | N-despropyl ropinirole | Mephenytoin ³ |
| Meprobamace | | 219→158 | 34 | | N-Acetylserotonin ¹ |
| Morphine | 20 | 286→152 | 5 | Norcodeine Hydromorphone Norhydrocodone Letrozole | 7-Aminoclonazepam ¹ Cladribine ¹ fludarabine ¹ |
| | | 286→165 | 6 | Isothipendyl N-Monodesmethyl-rizatriptan Mepyramine | Faropenem ¹ Probenecid ² |
| | 25 | 234→56 | 8 | N,N-Didesmethyltramadol | Dopamine 3/4-O-sulfate ³ |
| Normeperdine | | 234 > 42 | 5 | | Lomustine ² p-Chlorobenzene sulfonyl urea |
| Topiramate | 2000 | 362→265 | 67 | Methylhydroxygliclazide 7-Hydroxygliclazide Alogliptin | Disopyramide ¹ Hexetidine ¹ |
| | | 362→207 | 66 | Methylergonovine nor-Levomethadyl acetate | Cefacetrile ² Noracymethadol ³ |
| Zalpidana | 10 | 308→235 | 45 | 2-oxobrimonidine Alcaftadine | Nitazoxanide ² |
| Zolpidem | | 308→92 | 13 | Ibopamine Hydroxyterbinafine | Glutathione ¹ Tolnaftate ¹ |

Excluded interferences

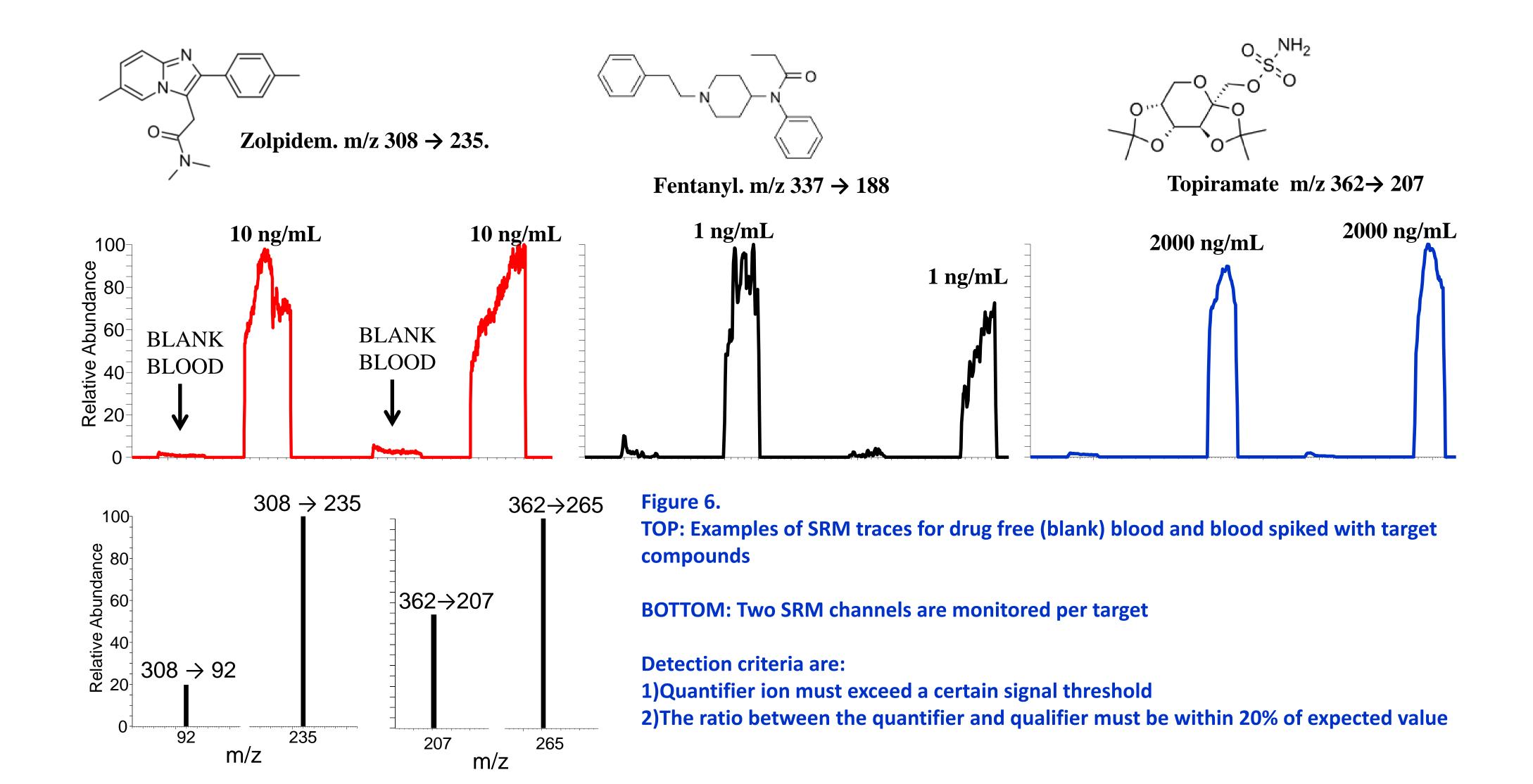
¹MS/MS spectra from the literature or database was compared to the target MS/MS spectra. There was no interference at the two target SRM channels. Main spectral source was Thermo's mzcloud

²compound is a poor ionizer and is unlikely to give appreciable MS signals

³compound is not available in the US and is also not an abused drug

Potential interferences

□Not enough information to exclude. Compounds will be tested for interferences if commercially available □The underlined compounds are known to interfere with the target compounds at one or both SRM channels



Conclusions

- ☐ From a subset of the most challenging drugs, paper spray MS/MS showed adequate sensitivity for 11 of the 14 targets for direct analysis of dried blood spots
- ☐ Only modest improvement is required to have adequate sensitivity for the other 3 targets.
- ☐ Within-class interferences, especially for opiates, is likely to occur
- ☐ No other source of false positives have been identified for this subset
- ☐ Next steps:
 - > Refine method to improve sensitivity (sample volume, extraction solvent)
 - Test remaining compounds to ensure adequate detection limits
 - > Use database searching to identify possible interfering compounds for the remaining target compounds. Experimentally test when needed.
 - ➤ Develop negative ion mode method (naproxen, salicylic acid, valproic acid, furosemide, hydrochlorothiazide, thiopental)

Acknowledgements

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